

Cryptosporidium spp.

Prevalence

- Infection of contemporary mouse colonies seldom reported (occasionally observed in wild rodent populations)
- Typically, not observed in healthy, conventional mice and rats
- Patent infection appears to be dependent on host intestinal microflora which assist resistance - suckling mice particularly at risk of **Cryptosporidiosis**
- SCID and athymic nude mice – unable to clear *C. muris* and *C. parvum* infections

Significance

- Clinically ill animals are not suitable for research
- Zoonotic risk to humans and potential danger to immunocompromised mice
- Suckling and immunosuppressed mice – useful in vivo model for anticryptosporidial compound testing
- Work done in immunocompromised models – elucidate immune mechanisms responsible for infection clearance

Disease

- *C. parvum* includes related species - *C. tyzzeri* (mouse genotype I) and mouse genotype II
- Clinical signs:
 - Infection may ascend biliary tract in nude and SCID mice – chronic cholangiohepatitis with focal hepatic coagulative necrosis
 - SCID mice infected with *C. parvum* and treated with dexamethasone – develop neoplasia in stomach, duodenum, and ileocecal region
 - Immunosuppressed – persistent infection with weight loss and sticky stools
 - Suckling mice infected – grow more slowly and are less active
 - Immunocompetent adult mice – develop transient infections
 - Heavy infection – mucosal hyperplasia and foci of dysplasia

Transmission

- Once consumed, sporozoites invade gastrointestinal epithelium and develop into merozoites
- Merozoites develop into micro- or macrogametes and eventually into sporulated oocysts
- Oocysts shed in faeces as fully infective parasites

Isolation and Diagnosis

- Histological examination of GIT tissue:
 - *C. muris* - infects glands of the gastric mucosa (relatively nonpathogenic):
 - gastric gland dilation and lymphoid infiltrate

- *C. parvum* (mice also susceptible to bovine genotype) – infects small intestinal epithelium (marginally pathogenic), heavy infections cause:
 - blunting and fusion of intestinal villi
 - crypt hyperplasia
 - lymphocytic-plasmacytic infiltration of underlying lamina propria
- *C. parvum* in athymic nude mice – hepatobiliary tree and pancreatic duct infection
- Routine hematoxylin-eosin staining of histological sections of the gastrointestinal mucosa
- Differences in size and tissue location – species differentiation
- Faecal floatation
- PCR

Prevention and Control

- Infection in immunocompetent animals and humans is self-limiting
- Not susceptible to chemotherapeutics
- Oocysts remain infective in chlorinated water and are resistant to common disinfectants
- Steam sterilization, pasteurization, and ethylene oxide exposure for 24hrs – inactivate oocysts
- Rederivation (through caesarean section or embryo transfer into pathogen free dams) and barrier maintenance

Reading

- S.W. Barthold, S.M. Griffey, & D.H. Percy. Pathology of Laboratory Rodents and Rabbits (Fourth Edition), 2016
- J.G. Fox, S.W. Barthold, M.T. Davisson, C.E. Newcomer, F.W. Quimby, A.L. Smith. The Mouse in Biomedical Research (Second Edition), 2007
- D.G. Baker. Flynn's Parasites of Laboratory Animals (Second Edition), 2007